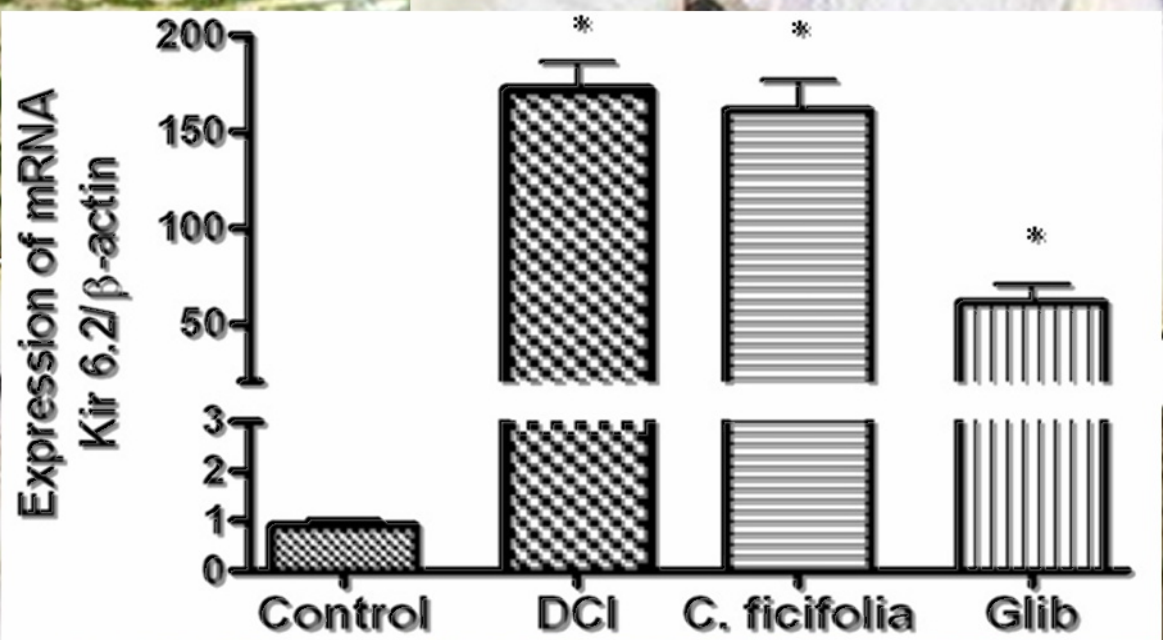


# International Biotechnology Color Journal

A Scientific Peer Reviewed Journal with Focus on BIOTECHNOLOGY  
and Covering Its Many Hues, Tints, Tones & Shades



Regular issue:

*Cucurbita ficifolia* Bouché fruit as insulin secretagogue.

Produced and hosted by *Centro de Investigación Científica de Yucatán, A.C.*,  
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Art, Science, Technology & Society*  
Int. biotechnol. color j., ISSN 2226-0404

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The International Biotechnology Color Journal appears every 4 months in the last week of February, June and October of each year, starting in October 2011. It is a non-profitable Electronic Publication. In its initial phase it does not require neither publication nor access fees. Its goal is the publication of Original Scientific Information, previously unpublished, and provided by the contributing authors out of its own will. *Int. biotechnol. color j.*, ISSN 2226-0404.

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## The INTERNATIONAL BIOTECHNOLOGY COLOR JOURNAL (IBCJ)

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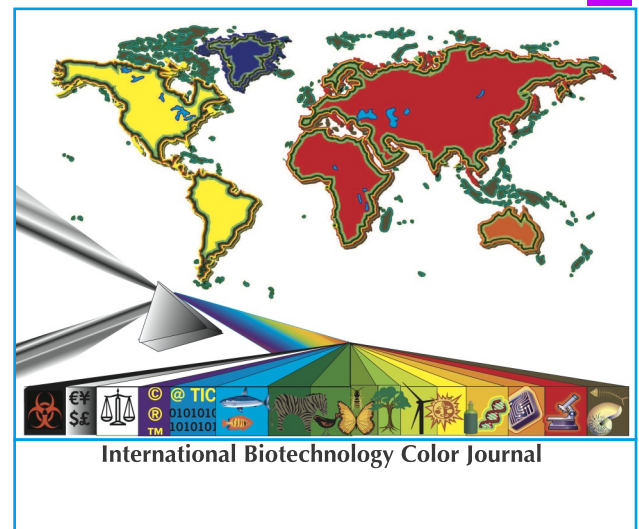
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Susana Lozano Muñiz  
President of the Foundation



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# International Biotechnology Color Journal

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## Editorial section

International Biotechnology Color Journal (IB CJ) is an electronic Open Access journal, devoted to the publication of peer-reviewed articles covering all the fields of biotechnology.

With this number we celebrate our first year of publication with main focus on the dissemination of scientific peer reviewed papers. In addition ICBJ intent is to provide a forum for reviews of special interest, notes presenting relevant findings in a short format, essays with new technical advances or relevant updates of reported protocols, book reviews, scientific meetings, and letters to the editor. The present issue offers a sample of such diversity.

Instructions for every type of contribution are presented in the journal's Homepage and at the end of the present issue. The Editorial Board of IBCJ is fully committed to publish novel contributions in all areas of biotechnology. Submissions are reviewed from a rigorous optic of scientific criticism and all original contributions within the scope of the journal are welcomed.

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## Editorial comments to the contents of this issue

By José Juan Zúñiga-Aguilar, Chief editor.

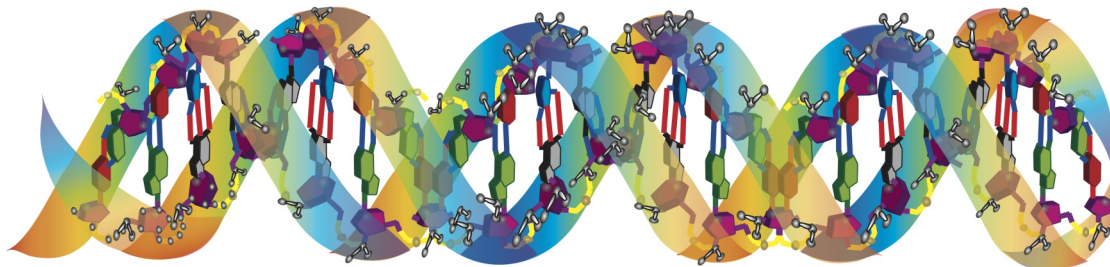
In this issue, a report of Miranda-Pérez *et al.* offers an interesting proposal on the mechanism hypoglycemic activity effects by extracts of *Cucurbita ficifolia* (chilacayote), an empiric effect largely accepted by the traditional knowledge. Using cultures of the insulin-producing RINm5F cells, they found that crude extracts of the plant, as well as samples of the D-chloro-inositol control treatment, promoted the increment of transcripts encoding insulin and Kir6.2 channels, respectively. This result offers relevant information supporting the traditional use of this plant as an alternative treatment of the type 2 Diabetes mellitus disease (T2D). Being T2D a public health problem in Mexico, the use of better-accepted drugs for its treatment constitutes an alternative to deal with a problem of high impact in health, economy and social development, and which is increasing dramatically in Mexico.

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Regular Issue, February 2013

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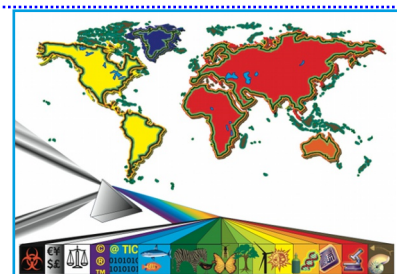
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## Why to publish on IBCJ?

By Rogelio Rodríguez-Sotres

Dear reader,

As the size of world's population increases, so does the scientific community. At the same time, technological aids arise in all areas and open the door to new opportunities. The editorials handling scientific communications are not an exception to this trend, and many new editorial houses have emerged.

Most of these new editorials operate under the Open Access Publishing model, commonly known as "author pays", contrary to the traditional publishing model where the reader covers the cost through his/her subscription fee.

The Open Access Model has grown rapidly because:

- A) Lowers production costs
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- C) Reduces "piracy" problems

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Unfortunately, as with any business, ethical misconduct happens, and some editorial may try to deceive authors setting up sites with all sorts of misleading data. These predatory editorials scam authors into sending their manuscripts with counterfeit invitations, and almost always, the contributors are not warned of the publications charges, until an invoice is issued.

There are very good editorials operating under the "author pays" model, and in fact, some of those editorials have created and published their own ethical guidelines, somehow setting a standard for the industry.

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A) We take our work seriously. The main staff of IBCJ are scientist, we understand the problems of the scientist, and specially those in the Latin America Scientific community, but we know scientist do not want their work to go published unreviewed and poorly edited.

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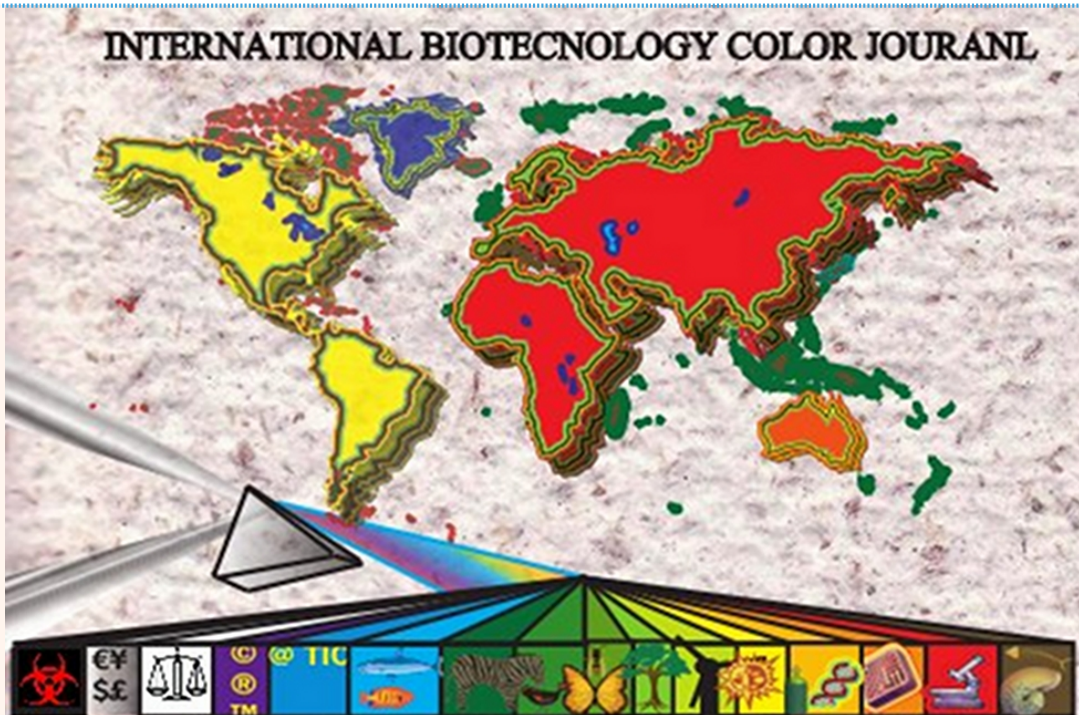
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# Cucurbita ficifolia Bouché fruit acts as an insulin secretagogue in RINm5F cells

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Chief Editor: Dr. José Juan Zúñiga Aguilar.

Received November 15<sup>th</sup>, 2012. Revised form February, 20<sup>th</sup>. Accepted February 28<sup>th</sup>.

## Abstract

Diabetes mellitus (DM) is an important global public health problem. Despite the availability of drug therapy, patients have continued using plants with anti-diabetic properties as alternative treatments for diabetes. *Cucurbita ficifolia* Bouché (*C. ficifolia*) is a plant cultivated in Mexico for its edible fruit that is medicinally used to control type 2 DM (T2D). The hypoglycemic effect of *C. ficifolia* has been demonstrated in different experimental models and in T2D patients. However, no studies have determined the mechanism of action of this hypoglycemic effect. The aim of the present investigation was to determine if the hypoglycemic action of *C. ficifolia* and D-chiro-Inositol (DCI; a compound found in the fruit of *C. ficifolia*) occurs through an increase in the production of insulin. An aqueous extract of this fruit was obtained and standardized by its content of DCI, the principal hypoglycemic compound of the fruit. To study the mechanism of hypoglycemic activity of this extract and DCI alone, RINm5F cells were exposed to different concentrations of both the extract and DCI, and the production of insulin and Kir6.2 channels were measured. The mRNA expression levels of insulin and Kir6.2 were found to be increased in cells that were treated with DCI and the *C. ficifolia* aqueous extract. This effect suggests a mechanism of action that involves both the expression and secretion of insulin. This research maintains the interest in developing new nutraceuticals from *C. ficifolia* fruits for use in the treatment of T2D.

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## Abbreviations

DM, diabetes mellitus; T2D, type 2 diabetes mellitus; SUR1, sulfonylurea receptor 1; DCI, D-chiro-inositol;

## Keywords

Type 2 diabetes, insulin, Kir6.2, *Cucurbita ficifolia* Bouché, D-chiro-inositol.



## Introduction

Type 2 diabetes mellitus (T2D) is a set of metabolic disorders characterized by increased levels of blood glucose (hyperglycemia) due to defects in insulin production and secretion by the pancreatic cells (1-4).

One of the key factors in the secretion of insulin by pancreatic cells is the ATP-dependent K<sup>+</sup> channels (K<sup>+</sup><sub>ATP</sub>). These channels are formed by two tetramers of the subfamily Kir6.2 and the sulfonylurea receptor 1 (SUR1) (5). Both Kir6.2 and SUR1 have the ability to induce changes in membrane electrical excitability. This change leads to calcium channel opening and an increase in cytoplasmic calcium concentration in beta cells, which triggers the mobilization and release of insulin (6). Sulfonylureas are anti-diabetic agents that act by releasing endogenous insulin following binding to SUR1 (7,8).

Despite the widespread use of antidiabetic drugs, more than 70% of the world population uses medicinal plants as the only available alternative for the treatment of their diabetes-related health problems (9). Furthermore, many medicinal plants have been reported to have "anti-diabetic" properties (10-12). *Cucurbita ficifolia* Bouché (*C. ficifolia*, Cucurbitaceae) is cultivated in Mexico for its edible fruit (13). It is an annual monoecious plant, which is grown primarily in the states of Mexico, Hidalgo, Puebla and Veracruz. *C. ficifolia* is commonly known as "chilacayote", and its immature fruit is used for preparing different dishes. The mature fruit of *C. ficifolia* is used to make a traditional candy (14). In the states of Mexico and Morelos, the annual production of *C. ficifolia* is 4,706 tons and the harvested area is 300 hectares (15).

*C. ficifolia* fruits are also medicinally used to control type 2 DM (T2D) and their hypoglycemic effect has been demonstrated in different experimental models and in T2D patients. D-chiro-inositol (DCI) has been proposed to be the active component in *C. ficifolia* (16). DCI, originally discovered as a mediator of intracellular insulin action, has been shown to accelerate the dephosphorylation of the glycogen synthase and pyruvate dehydrogenase, two regulatory enzymes of the glucose oxidation pathways (14,17). Clinical studies have demonstrated a linear relationship between body DCI deficiency and the degree of insulin resistance in subjects with T2D. In addition, the administration of DCI to diabetic rats and rhesus monkeys improved glucose utilization and insulin action (17). In streptozotocin-induced diabetic rats, daily administration of *C. ficifolia* fruit and DCI over a 30-day period reduced glycemia and increased the levels of liver glycogen, total hemoglobin and insulin (16). Although both *C. ficifolia* fruit extracts and DCI might be used as coadjuvants in DM control (18,19), there are no studies that explain the mechanisms of action involved in their hypoglycemic effect. This is very important because the juice of the fruit has been studied in Mexico as a hypoglycemic agent in experimental animal models and in diabetic patients (19).

Therefore, the aim of this work was to determine if the hypoglycemic effect of the *C. ficifolia* fruit extracts and DCI may be explained by a rise in the expression levels of insulin and Kir6.2 in RINmF5 cells.

## Material and methods

### Plant material

Fresh mature fruits of *C. ficifolia* with a diameter of 18–20 cm and an approximate weight of 4 kg were gathered in the Acolman municipality, State of Mexico during April of 2011. The endocarp, free of seeds, was cut into thin slices and placed in a container at room temperature with constant aeration for dehydration. The dried material was ground in an electrical Wiley mill, using a grid of 1 mm diameter (14). This material (100 g) was macerated with water (1 L) for 72 h in a laminar flow hood. The aqueous phase was filtered and centrifuged at 805 × g to obtain a precipitate, which was separated and freeze dried (LABCONCO) (yield = 5%) (19). Quantification of the DCI content in the *C. ficifolia* extract was performed using high performance liquid chromatography (HPLC Waters 2695 separation module) as reported by Roman Ramos et al. (2012) (14). This resulted in an extract with a concentration of 3.3 mg of DCI/g of extract. A pure DCI standard was obtained from Sigma-Aldrich.

### RINm5F cell culture

RINm5F, which is an insulin-producing cell line derived from a pancreatic islet tumor, cells were commercially acquired from the American Type Culture Collection (ATCC). RINm5F cells were grown in a monolayer culture using RPMI 1640 medium (11.1 mM glucose) (GIBCO™), supplemented with 10% fetal bovine serum (ATCC), 2 mM L-glutamine, 1 mM sodium pyruvate and 2 µg L<sup>-1</sup> of gentamycin (Invitrogen). The cells were grown at 37 °C in disposable plastic bottles (Nunc™) and in a humidified atmosphere of 5% CO<sub>2</sub>/95% air (20–22). The medium was replaced twice a week.

### MTT assay

Cell viability, following treatment with both the extract of *C. ficifolia* fruit and DCI alone, was measured using the 3-(4,5-dimethylthiazole-2-yl)-2,5-dihenyttetrazolium bromide (MTT, sigma) assay, according to Mosmann (23). The assay measures the conversion of MTT to insoluble

formazan by the dehydrogenase enzyme activity of the intact cells; this assay is widely used for studies of cytotoxicity, viability and proliferation. The amount of the formazan produced is directly proportional to the number of living cells (22). The RINm5F cells were seeded into 96-well microplates at a semi-confluent density (5000 cells/well). After 24 h, the medium was replaced with complete medium containing concentrations of DCI ranging from 0.1 to 50  $\mu\text{M}$  in the aqueous extract of *C. ficifolia* fruit or the equivalent concentrations of DCI alone (Sigma). The cells were treated for 24 h. They were then washed with phosphate-buffered saline (PBS), pH 7.4, and a solution of 0.1 mg mL<sup>-1</sup> MTT in PBS (pH 7.5) was added. The cells were incubated for 3 h at 37 °C, followed by washing with PBS. Then, 200  $\mu\text{L}$  of 40 mM HCl (prepared in isopropanol) was added to each well for 15 min to solubilize the produced formazan. The OD was read at 570 nm (23). The data were expressed as the percentage of viable cells following treatment with the aqueous extract of *C. ficifolia* and DCI compared to the control cells.

Because the treatments with the aqueous extract of *C. ficifolia* fruit were based on its DCI content, we chose the equivalent concentrations for the DCI alone treatments of the RINm5f cells and quantify the insulin and Kir6.2 mRNA expression levels. Thus, the choice of the concentrations to treat the cells with the aqueous extract of *C. ficifolia* fruit and with DCI alone (0.25  $\mu\text{M}$  of DCI in both cases) preceded

the extraction of total RNA from the treated cells.

RNA isolation, reverse transcriptase polymerase chain reaction and real time PCR

For the extraction of total RNA, cells were seeded at  $1 \times 10^6$  cells (RINm5f) per treatment in 6-well culture dishes and incubated for 24 h. Both the aqueous extract of *C. ficifolia* fruit and the DCI treatments were performed at a concentration of 0.25  $\mu\text{M}$  and over 24 h. After treatment, the cells were washed with 1 mL ice-cold PBS and solubilized with 1 mL of TRIzol<sup>®</sup> reagent (Invitrogen). RNA was treated with chloroform, centrifuged at  $12,235 \times g$  for 15 min at 4 °C and precipitated with ethanol as described by Chomczynski (21, 25). RNA was extracted and re-dissolved in diethylpyrocarbonate treated water and the OD at 260 nm was measured to determine the RNA concentration. cDNA was synthesized using 2  $\mu\text{g}$  total RNA by reverse transcriptase PCR using the ImProm II reverse transcription system (Promega). The cDNA was amplified by an enzyme DNA polymerase kit, the DNA master plus SYBR Green 1 PCR MasterMix (Applied Biosystems) (20) for the following genes: Insulin NM\_09129.3 (5' - TGCCAGGCTTTTGTCAAAC-3'), Kir6.2 NM\_031358.3 (5' -GTACAGATCATTGTGGCGT-3') and  $\beta$ -actin NM\_031144.3 (5' -GTGGGTATGGGTCAGAAGGA-3'), which was used as a control.

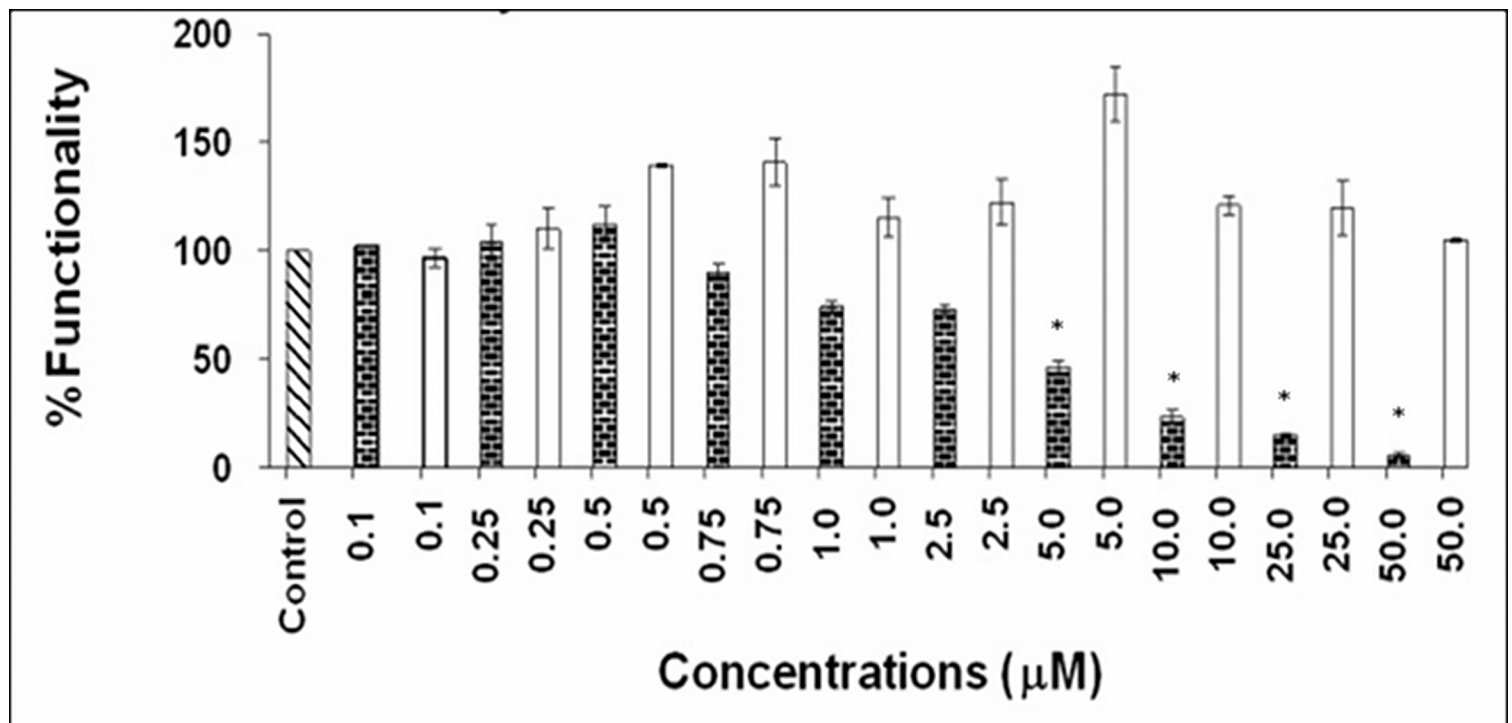


Figure 1. Function of *C. ficifolia* fruit extract and DCI in RINm5F cells. Cells were treated with the aqueous extract of *C. ficifolia* fruit and DCI at concentrations ranging from 0.1 to 50  $\mu\text{M}$  (x-axis). The Y-axis shows the percentage of activity in the treated cells. A concentration of 0.25  $\mu\text{M}$  was chosen for treating the cells (n=6). \*Significantly different compared to control (p<0.05).

## Statistical analysis

Data are presented as the mean  $\pm$  standard error of the mean (S.E.M.). Significant differences among the treatments were determined by an analysis of variance using the Tukey-Kramer Multiple Comparison post-hoc test ( $p < 0.05$ ).

## Results and Discussion

Fig. 1 shows the percentage of dehydrogenase functionality in the RINm5f cells treated with different concentrations of both the aqueous extract of *C. ficifolia* fruit and DCI. At a concentration of 1  $\mu$ M of DCI in the aqueous extract of *C. ficifolia* fruit, a 20% cell functionality decrease compared to the control group ( $p < 0.05$ ) was observed; this effect was indicative of a concentration-dependent cytotoxic effect. A DCI alone concentration of 0.25  $\mu$ M showed a retention of 98% of the dehydrogenase functionality of the cell compared to the control group. DCI alone did not show a cytotoxic effect at any of the concentrations examined. Therefore, a concentration of 0.25  $\mu$ M DCI was chosen to examine insulin and Kir6.2 gene expression following treatment with the aqueous extract of *C. ficifolia* fruit and DCI alone.

A statistically significant increase in insulin mRNA expression (a 5-fold increase) compared with the control group was observed after treatment with the aqueous extract of *C. ficifolia* fruit and with DCI alone in the RINm5F cells (Figure

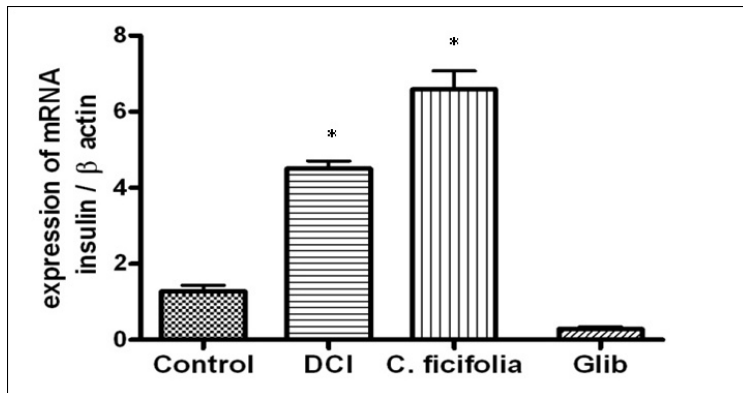


Figure 2. Insulin mRNA gene expression in RINm5F cells. Cells were treated with the aqueous extract of *C. ficifolia* fruit and DCI at a concentration of 0.25  $\mu$ M for 24 h. The aqueous extract of *C. ficifolia* fruit and DCI induced insulin gene expression, which was increased 5-fold ( $n=6$ ). \*Significantly different against control ( $p < 0.05$ ).

2). Therefore, the hypoglycemic effect of *C. ficifolia* may be explained by an increase in the expression of the insulin gene. Various studies that focused on gene therapy for diabetes have proposed the use of agents with a strong relationship between the stimulation of insulin gene expression and its secretion (26). Ultra-structural analysis of beta cells has shown reduced insulin granules in human D2T. These survival and functional changes are accompanied by modifications of beta cells gene and protein expression (27). Finding agents that regulate insulin gene expression to give an advantage to beta cells holds considerable promise and could provide the beta cells the necessary flexibility to adapt to the changing environment in DT2. If *C. ficifolia* exerts any effect on the regulation of gene expression of insulin, it would be a viable therapeutic agent for the treatment of T2D.

Studies in rat beta cells have associated the effect of glibenclamide on insulin mRNA expression with an increase in the cytoplasmic concentration of  $[Ca^{2+}]$  after 24 h treatment (28). Therefore, the aqueous extract of *C. ficifolia* fruit could stimulate insulin gene expression through an increase in cytoplasm  $[Ca^{2+}]$ . Although our results are in agreement with other studies in which *C. ficifolia* fruit extracts and DCI have shown an effect on insulin secretion (16), other studies are required that measure insulin secretion and the cytoplasmic concentration of  $[Ca^{2+}]$ . Nevertheless, in our studies, glibenclamide (24 h, 4  $\mu$ M) did not cause an increase in the expression of insulin mRNA as has been reported in previous studies (28). This is likely due to the difference in the treatment concentrations (24 h, 0.5  $\mu$ M).

ATP-sensitive potassium channels ( $K^+_{ATP}$ ) in pancreatic beta cells contain SUR1 and the inwardly rectifying potassium channel (Kir) 6.2 subunits. SUR1 and the Kir6.2 protein assemble into a potassium channel and play a key role in the regulation of insulin secretion (29). We evaluated the effect of the aqueous extract of *C. ficifolia* fruit and DCI on Kir6.2 mRNA expression. Figure 3 shows that in RINm5F cells, the aqueous extract of *C. ficifolia* fruit and DCI significantly increased Kir6.2 mRNA expression (greater than 100-fold) compared with the control group. This is the first report of the effect of DCI and the aqueous extract of *C. ficifolia* fruit on Kir6.2 mRNA expression. The  $K^+_{ATP}$  channel plays an important role in insulin secretion, and this investigation opens the possibility that in the hypoglycemic effect of DCI and *C. ficifolia* is involved the expression of both insulin and DCI. In other studies, the expression of Kir6.2 has been positively correlated with cellular protein content (30). Thus, the increase in Kir6.2 mRNA by the aqueous extract of *C. ficifolia* fruit and DCI should indicate an increase in the protein content, which would be crucial for the secretion of insulin. Other studies are required to confirm the association among Kir6.2 expression, protein content and insulin secretion.

Notably, the mechanism of action of glibenclamide involves a block of the  $K^+_{ATP}$  channel by binding to SUR1 in

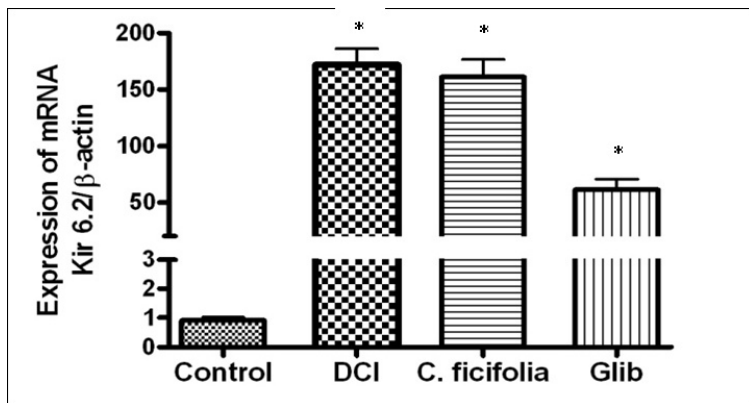


Figure 3. Kir 6.2 mRNA gene expression in RINm5F cells. Cells were treated with the aqueous extract of *C. ficifolia* fruit and DCI at a concentration of 0.25  $\mu$ M for 24 h. The aqueous extract of *C. ficifolia* fruit and DCI induced Kir6.2 gene expression, which was increased 100-fold (n=6). \*Significantly different against control ( $p < 0.05$ ).

beta cells. This depolarizes the cell plasma membrane, inducing the opening of  $[Ca^{2+}]$  channels and the secretion of insulin (31). In this study, glibenclamide stimulated the expression of Kir6.2 without affecting insulin expression (Figure 3). However, the *C. ficifolia* fruit extract and DCI increased the mRNA expression of both insulin and Kir6.2; therefore, the *C. ficifolia* fruit extract and DCI must have a different hypoglycemia inducing mechanism than glibenclamide. This is likely the result of distinct chemical components in the *C. ficifolia* fruit extract other than DCI.

Previous phytochemical studies have shown the presence of active principles in the aqueous extracts of *C. ficifolia* fruit (32). The aqueous extract contains DCI (3.3 mg/g of extract), and it has been reported to contain flavonoids and cucurbitacins. There have been similar findings in other Cucurbitacea species (33). Flavonoids are secondary metabolites of plants, and have been characterized as being soluble in water. Flavonoids have been principally associated with increased insulin secretion (34-35). Flavonoids are likely to be associated with the expression of insulin mRNA, which clearly agrees with the greater effect of the aqueous extract of the *C. ficifolia* fruit compared to DCI alone.

The fruit of *C. ficifolia* has been associated with a hypoglycemic effect, and it has been shown to have additional benefits as an antioxidant and an anti-inflammatory (10,14,19). It is probable that these actions may be attributed components in the extract other than DCI, such as, flavonoids (36) or cucurbitacins (37). However, a complete characterization of the aqueous extract is necessary to correlate these effects.

*C. ficifolia* has been reported as a useful alternative for the treatment of T2D (10-12,14). The activity of *C. ficifolia* and DCI on the mRNA expression levels of proteins implicated in

the production and secretion of insulin in pancreatic beta cells represents a notable finding. Other studies have focused on verifying the hypoglycemic effect of *C. ficifolia* in both diabetic patients and animal models (16,18-19). These studies have provided a great contribution to the study of medicinal plants. However, this study represents the first attempt to elucidate the mechanism of action of *C. ficifolia*: an increase in the expression of insulin and  $K^+_{ATP}$  channels, which is likely due to DCI.

Several nutraceuticals used in clinical practice have been shown to target the pathogenesis of diabetes mellitus (38). In our laboratory, a jelly developed from pomegranate juice with the addition of a pomegranate rind extract, has the potential to control DM (39). Therefore, in the same manner, we can develop a nutraceutical from *C. ficifolia*.

## Conclusion

The aqueous extract of *C. ficifolia* fruit and DCI increased mRNA expression for insulin and Kir6.2 in RINm5F cells. These results suggest a mechanism of action involving the stimulation of insulin and Kir6.2 (a protein widely involved in the secretion of insulin) production, which represents a useful alternative for DM control. Therefore, the fruits of *C. ficifolia* may be considered to be a potential source of raw material for obtaining new oral hypoglycemic drugs and for the development of a new nutraceutical for DM control.

## Acknowledgements

We thank CONACYT for the grants awarded to M.B.E Maria Elizabeth Miranda Perez. This research was partially supported by PROMEP-SEP (P/CA-15-2006-35-53). We appreciate the support of Dr. Maria Concepcion Gutierrez Ruiz, from the Laboratory of Cellular Physiology. The authors would also like to acknowledge Biol. Gerardo Garcia Velazquez, MSc Jesus Vladimir Hernandez-Rosado and MSc Jessica Garcia Gonzalez from the Postgrad in Experimental Biology at Universidad Autónoma Metropolitana, for their technical assistance.

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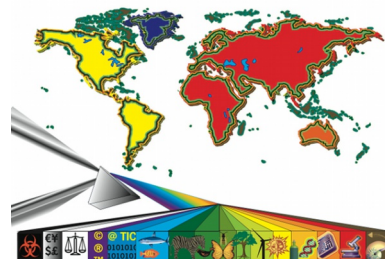


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